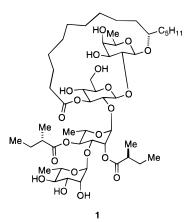
## Synthesis of the Macrolactone **Disaccharide Subunit of Tricolorin A**

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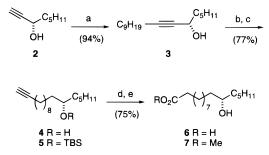
Farmers in the southeastern intertropical Mexican state of Morelos make extensive use of *Ipomoea tricolor* as a cover crop during the fallow period in sugar-cane fields. This tuberous plant has the useful property of suppressing the growth of other plants, including invasive weeds. In 1993, Pereda-Miranda and co-workers<sup>1</sup> reported the isolation of tricolorin A (1), the actual compound responsible for the biological activity of this plant. Compound 1 also demonstrated significant cytotoxicity against cultured P-388 and human breast cancer cells. We chose tricolorin A as a synthetic target because of the unique challenge in forming the macrolactone in this molecule. We now report the synthesis of a protected fucosyl  $\beta$ -D-glucoside that incorporates the 19-membered lactone characteristic of tricolorin A.

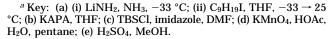


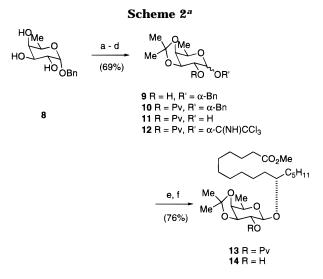
The synthesis of the hydroxy acid aglycone is summarized in Scheme 1. The (S)-propargylic alcohol  $2^2$  was deprotonated with LiNH<sub>2</sub> and the resulting lithioalkyne treated with 1-iodononane to obtain 3 in 94% yield. Treatment of 3 with KNH(CH<sub>2</sub>)<sub>3</sub>NH<sub>2</sub> (KAPA)<sup>3</sup> provided terminal alkyne 4 in 79% yield. Protection of the alcohol with tert-butyldimethylsilyl chloride, followed by oxidative cleavage of the alkyne to the corresponding acid,<sup>4</sup> and subsequent Fisher esterification gave the methyl ester. Additionally, the acidic esterification reaction conditions conveniently cleaved the TBS ether to give the desired hydroxy ester 7 in good overall yield.

The fucose unit was prepared from the known benzyl  $\alpha$ -D-fucopyranoside (Scheme 2).<sup>5</sup> Selective protection of the C-3 and C-4 hydroxyl groups as the acetonide followed by protection of the C-2 hydroxyl group as the pivaloyl ester gave the fully protected intermediate 10. The anomeric position was then unmasked by catalytic

Scheme 1<sup>a</sup>







<sup>a</sup> Key: (a) 2,2-dimethoxypropane, p-TsOH; (b) t-BuCOCl, pyridine, DMAP, 70 °C; (c) 50 psi H<sub>2</sub>, Pd(OH)<sub>2</sub>, EtOAc; (d) Cl<sub>3</sub>CCN, Cs<sub>2</sub>CO<sub>3</sub>, CH<sub>2</sub>Cl<sub>2</sub>; (e) 7, TMSOTf, CH<sub>2</sub>Cl<sub>2</sub>; (f) NaOMe, MeOH, MeOAc.

hydrogenation of the benzyl ether. Activation of the fucose derivative for glycoside formation was achieved by treatment with Cl<sub>3</sub>CCN and Cs<sub>2</sub>CO<sub>3</sub><sup>6</sup> to furnish trichloroacetimidate 12.

Coupling of hydroxy ester 7 and the crude trichloroacetimidate occurred smoothly in CH<sub>2</sub>Cl<sub>2</sub> with catalytic TMSOTf<sup>7</sup> to give the desired  $\beta$ -glycosidic linkage in 79% yield. The C-2 hydroxyl group was exposed by cleavage of the pivaloyl ester with NaOMe in a MeOH/MeOAc cosolvent to give coupling partner 14. A large excess of NaOMe was employed in this reaction to allow the reaction to proceed at a practical rate. We found that use of MeOAc in this step greatly minimized saponification of the methyl ester functionality in the molecule by a minor amount of hydroxide present in the NaOMe reaction solution.

As shown in Scheme 3, the glucose unit preparation began by protection of the known glucopyranose 15<sup>8</sup> to form the triacetyl compound 16. Formation of the amino glycoside by treatment with BnNH<sub>2</sub> followed by selective hydrolysis with dilute aqueous acid<sup>9</sup> furnished pyranose

<sup>(1)</sup> Pereda-Miranda, R.; Mata, R.; Anaya, A. L.; Wickramaratne, D.

<sup>B. M.; Pezzuto, J. M.; Kinghorn, A. D. J. Nat. Prod. 1993, 56, 571.
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<sup>(4)</sup> Krapcho, A. P.; Larson, J. R.; Eldridge, J. M. J. Org. Chem. 1977, 42. 3749.

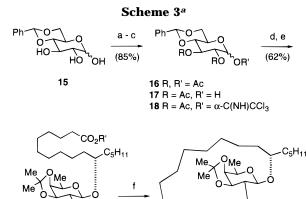
<sup>(5)</sup> Heyns, K.; Baron, A. L.; Paulsen, H. Chem. Ber. 1964, 97, 921.

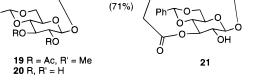
<sup>(6)</sup> Urban, F. J.; Moore, B. S.; Breitenbach, R. Tetrahedron Lett. 1990, 31, 4421.

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Ph





 $^a$  Key: (a) Ac\_2O, Et\_3N, DMAP, CH\_2Cl\_2; (b) (i) BnNH\_2, THF, (ii) 1 N HCl; (c) Cl\_3CCN, Cs\_2CO\_3, CH\_2Cl\_2; (d) **14**, AgOTf, CH\_2Cl\_2; (e) LiOH, THF, H\_2O; (f) 2,4,6-trichlorobenzoyl chloride, Et\_3N, DMAP, benzene.

17 in 87% yield.<sup>10</sup> Glycosyl donor 18 was obtained by treatment of 17 with  $Cl_3CCN$  and  $Cs_2CO_3$ .

Treatment of alcohol **14** with the crude trichloroacetimidate and anhydrous AgOTf<sup>11</sup> in CH<sub>2</sub>Cl<sub>2</sub> gave an 84% yield of the  $\beta$ -disaccharide. Simultaneous saponification of the three ester groups in disaccharide **19** with LiOH gave the macrolactonization precursor **20** in good yield. Following the Yonemitsu protocol,<sup>12</sup> the acid diol lactonized at the C-3 hydroxyl position of the glucose ring with a high degree of selectivity over the C-2 position to give the target lactone **21** in 71% yield.

In summary, the target lactone disaccharide has been synthesized in a total of 17 steps, with a longest linear sequence of 10 steps and an overall yield of 18%. We are currently exploring the synthesis of a suitable rhamnosyl  $\alpha$ -L-rhamnopyranoside to use as a glycosyl donor for converting **21** into tricolorin A.

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**Supporting Information Available:** Experimental procedures and spectral data for all compounds (9 pages).

## JO960655B

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<sup>(10)</sup> Compounds **16** and **17** have been previously reported in the literature, but were prepared by different methods and no modern spectral data is available. For compound **16**, see: Zervas, L. *Chem. Ber.* **1931**, *64*, 2289. For compound **17**, see: Korytnyk, W.; Mills, J. A. *J. Chem. Soc.* **1959**, 636.

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